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Frequencies of Genetic Polymorphisms of the Cholesterol and Statin Metabolic Pathway Genes among Healthy South Indian Tamil Population

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ABSTRACT

Background: Indians are genetically predisposed to Coronary Artery Disease (CAD), hence it is worth studying the effect of genetic polymorphisms which affect CAD development including Low- Density Lipoprotein Cholesterol (LDL-C) in our population. Studies have found that there is a considerable variation in the genetic makeup of Indians with distinct genetic groups having been identified-one such being Dravidian Tamil population. **Aim:**We aimed to determine the distribution of allele and genotype frequencies of genes associated with LDL-C lipid levels as well as those associated with statin metabolic pathway in 100 healthy South Indian Tamil volunteers. **Results:**The minor allele frequencies (MAFs) of the genetic polymorphisms of *HMGCR* rs5908, rs17238540 and rs12916 were 0.5, 3.5 and 41% respectively. The MAF of *LDLR* rs688, *CYP7A1* rs3808607, ABCB1 rs1128503, *SLCO1B1* rs4149056 were 26, 42, 41.5 and 7% respectively. The frequencies of the genetic polymorphisms studied show considerable variation from other Indian ethnic groups in 5 out of the 7 genetic poly-

morphisms studied. **Conclusion:** Considerable variations in frequencies of genetic polymorphisms were found between Tamil population and other major ethnic populations worldwide with respect to *HMGCR* rs17238540 and *LDLR* rs688.

Key words: Genetic polymorphisms, Genotype frequency, Tamilians, Healthy volunteers, LDL-C, Statin pathway.

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INTRODUCTION

Elevated levels of Low-Density Lipoprotein Cholesterol (LDL-C) is associated with an increased risk of coronary artery disease (CAD).¹ LDL-C levels are affected by genetic and environmental factors. The basal risk in the form of genetic variations is not modifiable, although they are to a good extent modulated by environmental factors. Exploring and characterizing these genetic variations for each population would provide the background information needed to conduct further studies on lipid abnormalities and their treatment. Lipid traits are up to 60% heritable.² Indians being predisposed to CAD,³ it is essential to study the genetics of LDL-C to help predict CAD and to implement preventive measures in this high- risk population. Till now, 157 gene loci have been studied for association with blood lipid levels which could explain only 12-14% variation in the lipid levels.⁴

The clinical significance of genetic variants associated with lipid abnormalities depends on the frequency of genetic variants in the population of interest. Further, the normative data of frequency of single nucleotide polymorphism (SNP) is essential for planning and conducting genotypephenotype association studies. The frequency distribution of SNPs in other population cannot be used for planning studies in Indian population due to their unique genetic constitution.⁵

In earlier studies, the frequencies of genetic polymorphisms of genes involved in lipid and statin pathways such as *ApoA*, *ApoB*,⁶ *CETP*,⁷ ABCB1,^{8,9} and *CYP3A5*¹⁰ have been established among healthy South Indian Tamil population in our laboratory. For the current study, SNPs

were selected based on their role in statin and cholesterol metabolism. These variants have been associated with altered LDL-C response to statins or associated with toxicity in other populations. However, there is no data among South Indian Tamil patients. Genes such as HMGCR, LDLR and CYP7A1 are involved in the cholesterol metabolic pathway. HMGCR codes for 3-hydroxy-3-methylglutaryl-CoA reductase which is the rate-limiting enzyme for cholesterol synthesis.11 LDLR codes for low -density lipoprotein receptor, which is involved in endocytosis of LDL cholesterol.¹² CYP7A1 belongs to Cytochrome 450 group of enzymes which catalyzes the rate limiting step in cholesterol catabolism to bile acids, the major route of cholesterol elimination from the body.¹³ ABCB1 encodes the membrane associated protein which is a member of the superfamily of ATP-binding cassette (ABC) transporters which is involved in efflux of statins.14 The SLCO1B1 codes for hepatic solute carrier organic anion transporter family member 1B1 which is responsible for uptake of bilirubin and its metabolites and drugs like statins.¹⁵

Hence in the current study, we aimed to determine the frequencies of genetic variations for other genes in the lipid pathway such as *HMGCR* (rs5908, rs17238540, rs12916), *LDLR* (rs688), *CYP7A1* (rs3808607), *ABCB1* (rs1128503), and *SLCO1B1* (rs4149056) in South Indian Tamil population. We also aimed to compare the studied genotype distributions with that of other ethnic populations.

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MATERIALS AND METHODS

The study was approved by the Institute Ethics Committee (Human Studies) of Jawaharlal Institute of Postgraduate Medical Education and Research (JIPMER), Puducherry, India. Written informed consent was obtained from participants recruited for the study. The study was conducted among 100 apparently healthy volunteers of either gender, aged between 27 and 70 years. All study subjects belonged to South Indian Tamil ethnicity which was defined as people with history of their past three generations residing in the state of Tamil Nadu or Puducherry and speaking Tamil language as the mother tongue.

Five milliliters of blood for genotyping was collected from the forearm of the patient in sitting posture, using ethylenediamine tetra-acetic acid (EDTA) as the anticoagulant. The cellular layer including the buffy coat was separated using ultracentrifugation at 2500 rotations per minute (RPM) for 10 min, and stored at -20° C, until extraction. DNA extraction was performed using standard phenol chloroform method. Genotyping was done with allelic discrimination assays, with the kits obtained from Applied BioSystems (ABI) USA. The assay kits were based on TaqMan Technology. Real-Time PCR platform ABI 7300 was used. The allelic call was read with the help of sequence manipulation suite (SMS) version 1.4. The genotyping results were confirmed by running the samples in triplicates (representative genotypes) and by Sanger sequencing.

Hardy-Weinberg equilibrium for the studied genotypes was tested using Chi-square test. Genotype frequencies from the current study were compared with data from other studies using Fisher's Exact Test or Chisquare test. P value less than 0.05 was considered significant. All statistical tests were done using GraphPad Instat v 3.0 (GraphPad Software Inc., San Diego, CA, USA). study are given in Table 1. *HMGCR rs5908* was found to be very rare in the study population with only one subject having a heterozygous variant genotype with the absence of homozygous variant genotype. *HMGCR* rs17238540 polymorphism was also less prevalent in our population with the frequency of 3.5%. The variant alleles of *HMGCR* rs12916, *CY-P7A1* rs3808607 and *ABCB1* rs1128503 polymorphisms were found to be more prevalent. The variant genotype of *SLCO1B1* rs4149056 polymorphism was not found in our population. All genotypes except *LDLR* rs688 polymorphism did not show significant deviation from Hardy -Weinberg equilibrium.

Data of genotype and allele counts of the studied SNPs from North India¹⁶ and other major ethnic groups retrieved from 1000 genomes project phase 3 population¹⁷ are given in Table 2. The distribution of variant genotypes of HMGCR rs5908 observed in our population was significantly different only from North Indian population. However, distribution of HMGCR rs17238540 genotypes were significantly different from other populations except for CEU (Utah residents with Northern and Western European Ancestry), MXL (Mexican Ancestry in Los-Angeles, California), and Europeans from Norfolk-United Kingdom.¹⁸ The distribution of HMGCR rs12916 genotypes was significantly different from that of CEU, YRI (Yoruba in Ibadan, Nigeria) and MXL population. The distribution of LDLR rs688 genotypes in the current study was distinctly different from the other populations except that of CHB (Han Chinese in Beijing, China). The distribution of genotypes of CYP7A1 rs3808607 polymorphism was significantly different from a north Indian study which included patients from Punjab, Haryana and Chandigarh and also from YRI and JPT (Japanese in Tokyo, Japan)but not from other population of North India.¹⁹ The distribution of ABCB1 rs1128503 was significantly different from that of CEU and YRI population. The distribution of genotypes of SLCO1B1 in our population was significantly different from that of GIH (Gujarati Indian in Houston), CEU, CHB, and YRI.

RESULTS

The study subjects included 75 females and 25 males with a mean age of 45.5 ± 10.9 (SD). The genotype and allele frequencies established in this

Table 1: Genotype and allelic frequencies of the studied genetic polymorphisms

Gene	SNP	C	Genotype (%	b)	Allele (%)		HWE p-value	
	1912 A>G (rs5908)	AA	AG	GG	Α	G		
	1912 A/G (185906)	99	1	0	99.5	0.5	0.95	
HMGCR	74655498T>G (rs17238540)	TT	TG	GG	Т	G		
		93	7	0	96.5	3.5	0.71	
	*372C>T (rs129 16)	CC	CT	TT	С	Т		
		36	46	18	59	41	0.62	
LDLR	1773 C>T (rs688)	CC	СТ	TT	С	Т		
LDLK		64	20	16	74	26	< 0.0001*	
OVD741	505002(5C) T (GG	GT	GT TT G T				
CYP7A1	58500365G>T (rs3808607)	22	40	38	42	58	0.07	
	97550295 A. C (==1129502)	GG	AG	AG AA G A				
ABCB1	87550285A>G (rs1128503)	18	47	35	41.5	58.5	0.74	
	521 T>C (rs4149056)	TT	СТ	CC	Т	С		
SLCO1B1		86	14	0	93	7	0.45	

HWE - Hardy Weinberg Equilibrium p value.

*p value less than 0.05, significant deviation from Hardy Weinberg Equilibrium

SNP	Population	N	Gen	Genotype count			count	P value
HMGCR rs5908	Tamilians	100	AA	AG	GG	A G		
			99	(1	0)	199	1	
	North Indians[16]	150	126	(22	2)	274	26	< 0.0001
	GIH[17]	103	103	0	0	206	0	0.49
	ITU#	102	102	0	0	204	0	0.49
	BEB#	86	85	(1	0)	171	1	1.00
	PJL#	96	93	(3	0)	189	3	0.36
	STU#	102	100	(2	0)	202	2	1.00
	CEU#	99	94	(5	0)	193	5	0.12
	CHB#	103	103	0	0	206	0	0.49
	YRI#	108	108	0	0	216	0	0.48
	JPT#	104	104	0	0	208	0	0.49
	MXL#	64	63	(1	0)	127	1	1.00
HMGCR	Tamilians	100	тт	GT	GG	т	G	
rs17238540		100	93	(7	0)	193	7	
	North Indians	150	150	0	0	300	0	0.001*
	GIH	103	103	0	0	206	0	0.006*
	ITU	102	102	0	0	204	0	0.006*
	BEB	86	86	0	0	172	0	0.01*
	PJL	96	96	0	0	192	0	0.01*
	STU	102	101	(1	0)	203	1	0.03*
	CEU	99	96	(3	0)	195	3	0.33
	CHB	103	103	0	0	206	0	0.006*
	YRI	108	88	(19	1)	195	21	0.02*
	JPT	104	104	0	0	208	0	0.006*
	MXL	64	59	(5	0)	123	5	1.00
	European[18]	23011	22010	(989	12)	45009	1013	0.19
HMGCR rs12916	Tamilians	100	сс	ст	тт	с	т	
			36	46	18	118	82	
	North Indians	150	64	63	23	191	109	0.56
	GIH	103	35	55	13	125	81	0.45
	ITU	102	33	39	30	105	99	0.15
	BEB	86	31	37	18	99	73	0.86
	PJL	96	25	52	19	102	90	0.31
	STU	102	33	49	20	115	89	0.85
	CEU	99	17	46	36	80	118	0.001*
	CHB	103	29	49	25	107	99	0.37
	YRI	108	7	28	73	42	174	< 0.0001
	JPT	104	26	58	20	110	98	0.22
	MXL	64	12	24	28	48	80	0.001*
	Tamilians	100	сс	ст	тт	с	т	
LDLR rs688		100	64	20	16	148	52	
	GIH	103	42	45	16	129	77	0.0009*
	ITU	102	35	52	15	122	82	< 0.0001
	BEB	86	38	36	12	112	60	0.004*

Table 2: Comparison of genotype counts of Tamilian population with other populations

SNP	Population	N	Genotype count			Allele count		P value
	STU	102	41	45	16	127	77	0.0007*
	CEU	99	35	42	22	112	86	0.0002*
	CHB	103	74	23	6	171	35	0.06
	YRI	108	105	3	0	213	3	< 0.0001*
	JPT	104	73	29	2	175	33	0.001*
	MXL	64	18	39	7	75	53	< 0.0001*
CYP7A1 rs3808607	Tamilians	100	GG	GT	тт	G	т	
		100	22	40	38	84	116	
	North Indians[19]	200	29	101	70	159	241	0.13
	North Indians[16]	150	8	38	104	54	246	< 0.0001*
	GIH	103	18	52	33	88	118	0.32
	ITU	102	23	52	27	98	106	0.18
	BEB	86	18	48	20	84	88	0.05
	PJL	96	13	42	41	68	124	0.30
	STU	102	27	46	29	100	104	0.34
	CEU	99	16	40	43	72	126	0.53
	CHB	103	23	44	36	90	116	0.89
	YRI	108	37	55	16	129	87	0.0006*
	JPT	104	30	53	21	113	95	0.01*
	MXL	64	5	31	28	41	87	0.056
ABCB1 rs1128503		100	GG	AG	AA	G	Α	
	Tamilians		18	47	35	83	117	
	GIH	103	17	55	31	89	117	0.65
	ITU	102	15	53	34	83	121	0.73
	BEB	86	12	40	34	64	108	0.69
	PJL	96	22	46	28	90	102	0.57
	STU	102	18	42	42	78	126	0.63
	CEU	99	29	55	15	113	85	0.003*
	CHB	103	11	40	52	62	144	0.06
	YRI	108	78	29	1	185	31	< 0.0001*
	JPT	104	18	47	39	83	125	0.93
	MXL	64	17	34	13	68	60	0.10
SLCO1B1			тт	ст	сс	т	с	
rs4149056	Tamilians	100	86	14	0	186	14	
	GIH	103	99	4	0	202	4	0.01*
	ITU	102	90	11	1	191	13	0.63
	BEB	86	77	9	0	163	9	0.46
	PJL	96	90	5	1	185	7	0.07
	STU	102	93	9	0	195	9	0.24
	CEU	99	71	27	1	169	29	0.01*
	СНВ	103	77	24	2	178	28	0.04*
	YRI	108	106	2	0	214	20	0.001*
	JPT	104	82	19	3	183	25	0.18
	MXL	64	54	10	0	118	10	0.77

Table 2: Comparison of genotype counts of Tamilian population with other populations

#Data obtained from 1000 genome project; GIH- Gujarati Indian in Houston, TX, ITU-Indian Telugu in the UK, BEB-Bengali in Bangladesh, PJL-Punjabi in Lahore, Pakistan, STU-Sri Lankan Tamil in the UK, CEU-Utah residents with Northern and Western European Ancestry, CHB-Han Chinese in Beijing, China, YRI-Yoruba in Ibadan, Nigeria, JPT-Japanese in Tokyo Japan, MXL-Mexican Ancestry in Los-Angeles, California. Parentheses () indicates genotypes were combined for analysis, due to low numbers. Genotype count from current study were compared with data from other studies using Fisher's Exact Test or Chi square test.

DISCUSSION

The present study has determined the normative data on allele and genotype frequencies for SNPs of five genes namely, *HMGCR* (rs5908, rs17238540, rs12916), *LDLR* (rs688), *CYP7A1* (rs3808607), *ABCB1* (rs1128503), and *SLCO1B1* (rs4149056). These genes are involved in the maintenance of plasma lipid levels and the metabolism of statin drugs. To the best of our knowledge this study is the first to describe this information for South Indian Tamil population. The normative data on allele and genotype frequencies are important for planning of any genetic study to estimate sample size and to understand the clinical implications of a genetic variation. Thus, the current study provides the background information needed for future studies on pharmacogenetics of lipid metabolism and statins among South Indian Tamil population.

Genetic variants of HMG-CoA reductase (*HMGCR*) have been widely studied for their effect on lipid levels and variations in therapeutic responses to different statins. *HMGCR* variants have been associated with diminished response to pravastatin and simvastatin but not to fluvastatin²⁰ and atorvastatin.²¹ Further, they were also found to be associated with varied response between different ethnic groups.²² Among the three SNPs studied in *HMGCR* gene, variant genotypes of rs12916 and rs17238540 are more common in the study population. In contrast, *HMGCR* rs5908 variant being in low frequency may have less clinical relevance for the study population, even if functionally significant. Functional studies on rs12916 and rs17238540 variants may have clinical relevance in elucidating the effect of these SNPs on lipid homeostasis.

The HMGCR rs17238540 was found to be associated with reduced efficacy of statins in terms of lowering plasma levels of total cholesterol (TC) and LDL-C. Individuals with heterozygous variant genotype (GT) had 19% lower reduction in LDL-C when treated with pravastatin.²³ In the GoDARTS study, individuals with HMGCR rs17238540 variant G allele were associated with failure to achieve lipid lowering target.²⁴ In a study by Poduri et al., in North Indian population, the variant genotypes of HMGCR rs17238540 was associated with significantly higher LDL-C levels after atorvastatin therapy compared to wild type.²⁵ However, in a study by Thompson et al., this SNP was not associated with response to atorvastatin.²¹ In few studies, HMGCR rs17238540 was not associated with baseline lipid values.^{18,23,24} In the multi-ethnic study of atherosclerosis the HMGCR haplotypes,26 and these genotypes among North Indians were associated with baseline lipid levels respectively.²⁵ It was also observed that the HMGCR rs17238540 might negate some of the observed pleiotropic effects of statins, such as improved endothelial function, decreased platelet aggregability and reduced vascular inflammation and was also found to be associated with stroke risk in the EPIC-Norfolk study.27

The *HMGCR* rs12916 locus has been associated with total cholesterol and LDL-C levels among Europeans as well as East Asians and South Asians in a genome wide study.²⁸ In candidate gene studies the mean percentage reduction in LDL-C with *HMGCR* rs12916 polymorphisms among Chinese were found to be highest for homozygous variant, least for wild genotype and intermediate for heterozygous individuals after treatment with statins²⁹ while, studies among North Indians revealed that variant genotypes of this polymorphisms were responsible for poor response to atorvastatin in terms of LDL-C lowering.²⁵ Given the high prevalence of this genetic polymorphism among Tamilians, it would be of interest to determine the effect of this polymorphism on the lipid levels in our population.

LDL receptor gene (LDLR) is a commonly studied genetic locus for dyslipidemias. Genetic variations in LDL receptor gene may greatly reduce or abolish the function of LDL receptor. This may lead to increase in circulating LDL-C levels and risk of CAD.³⁰ Although LDLR gene

mutations are associated with familial hypercholesterolemia (FH), less dysfunctional variants have been associated with response to statin in non –FH individuals.³¹ Studies by Haiyan Zhu and his colleagues have identified SNP *LDLR* rs688 to be present in 60% of Caucasians and associated with a significant 10% increase in total and LDL-cholesterol in pre-menopausal women.³² This SNP has been recently shown to alter splicing efficiency, with the T allele being associated with increased total and LDL-cholesterol levels among premenopausal women.³³ Since this SNP is present in high frequency in the study population, and since it has not been studied among Indian population at large, more studies on the functional significance, of this SNP might yield useful information on the genetics of LDL-C in our population.

The genotype distribution of *CYP7A1* rs3808607 variants among Tamilian population was found to be similar to another study involving people from Lucknow region of North India,¹⁹ whereas it was significantly different from the results obtained in a study among North Indian regions of Punjab, Haryana, and Chandigarh,¹⁶ revealing the diversity even among North Indians. The genetic variants of *CYP7A1* rs3808607, *ABCB1* rs1128503, and *SLCO1B1* rs4149056, although similar to most other populations in frequency distribution, were found to occur at a higher frequency in the study population and may have clinical relevance if functional significance can be demonstrated in future studies.

CONCLUSION

The study has established the allele and genotype frequency distributions for *HMGCR* (rs5908, rs17238540, rs12916), *LDLR* (rs688), *CYP7A1* (rs3808607), *ABCB1* (rs1128503), and *SLCO1B1* (rs4149056) genetic variants among healthy subjects of South Indian Tamil population. The South Indian Tamil population is a unique ethnic group in terms of genetic polymorphisms in the cholesterol and statin metabolic pathways.

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CONFLICTS OF INTEREST

There are no conflicts of interest.

ABBREVIATION USED

ApoA: Apolipoprotein A; *ApoB*: Apolipoprotein B; *CETP*: Cholesteryl ester transfer protein; *ABCB1*: ATP binding Cassette subfamily B member 1; *CYP3A5*: Cytochrome P450 family 3 subfamily A member 5; *CY*-*P7A1*: Cytochrome P450 family 7 subfamily A member 1.

REFERENCES

- National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III). Third Report of the National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III) final report. Circulation. 2002;106(25):3143-421.
- Tada H, Won H-H, Melander O, Yang J, Peloso GM, Kathiresan S. Multiple associated variants increase the heritability explained for plasma lipids and coronary artery disease. Circ Cardiovasc Genet. 2014;7(5):583-7.
- 3. Enas EA, Senthilkumar A. Coronary Artery Disease in Asian Indians: An Update

And Review. The Internet Journal of Cardiology [Internet]. 2001 Dec 31 [cited 2017 Jan 11];1(2). Available from: http://ispub.com/IJC/1/2/4493.

- Justesen JM, Allin KH, Sandholt CH, Borglykke A, Krarup NT, Grarup N, et al. Interactions of Lipid Genetic Risk Scores With Estimates of Metabolic Health in a Danish Population. Circ Cardiovasc Genet. 2015;8(3):465-72.
- Reich D, Thangaraj K, Patterson N, Price AL, Singh L. Reconstructing Indian population history. Nature. 2009;461(7263):489-94. doi:10.1038/nature08365.
- Padmaja N, Ravindra KM, Adithan C. Apolipoprotein Al and Apolipoprotein B gene polymorphisms and lipid profile in Tamilian population. Ann Hum Biol. 2009;36(2):220-7
- Padmaja N, Ravindra KM, Soya SS, Adithan C. Common variants of Cholesteryl ester transfer protein gene and their association with lipid parameters in healthy volunteers of Tamilian population. Clinica Chimica Acta. 2007;375(1-2):140-6.
- Ramasamy K, Sisy Sam S, Chandrasekaran A. Allele and genotype frequency of MDR1 C3435T in Tamilian population. Drug Metab Pharmacokinet. 2006;21(6):506-8.
- Umamaheswaran G, Krishna KD, Kayathiri D, Rajan S, Shewade DG, Dkhar SA, et al. Inter and intra-ethnic differences in the distribution of the molecular variants of TPMT, UGT1A1 and MDR1 genes in the South Indian population. Mol Biol Rep. 2012 May;39(5):6343–51.
- Krishnakumar D, Gurusamy U, Dhandapani K, Surendiran A, Baghel R, Kukreti R, *et al.* Genetic polymorphisms of drug-metabolizing phase I enzymes CY-P2E1, CYP2A6 and CYP3A5 in South Indian population. Fundam Clin Pharmacol. 2012;26(2):295-06.
- 11. HMGCR Gene GeneCards | HMDH Protein | HMDH Antibody [Internet]. [cited 2016 Oct 25]. Available from: http://www.genecards.org/cgi-bin/carddisp. pl?gene=HMGCR.
- 12. LDLR Gene GeneCards | LDLR Protein | LDLR Antibody [Internet]. [cited 2016 Oct 25]. Available from: http://www.genecards.org/cgi-bin/carddisp. pl?gene=ldlr.
- CYP7A1 Gene GeneCards | CP7A1 Protein | CP7A1 Antibody [Internet]. [cited 2016 Oct 23]. Available from: http://www.genecards.org/cgi-bin/carddisp. pl?gene=CYP7A1.
- ABCB1 Gene GeneCards | MDR1 Protein | MDR1 Antibody [Internet]. [cited 2016 Oct 25].Availablefrom:http://www.genecards.org/cgi bin/carddisp. pl?gene=ABCB1&keywords=ABCB1.
- SLCO1B1 Gene GeneCards | SO1B1 Protein | SO1B1 Antibody [Internet]. [cited 2016 Oct 23].Availablefrom:http://www.genecards.org/cgi-bin/carddisp.pl ?gene=SLCO1B1&keywords=SLCO1B1.
- Poduri A, Khullar M, Bahl A, Sharma Y, Talwar K. A Combination of Proatherogenic Single-Nucleotide Polymorphisms Is Associated with Increased Risk of Coronary Artery Disease and Myocardial Infarction in Asian Indians. DNA and Cell Biology. 2009;28(9):451-60.
- A global reference for human genetic variation, The 1000 Genomes Project Consortium, Nature.2015 Oct; 526: 68-74. doi:10.1038/nature15393.
- Freitas RN, Khaw K-T, Wu K, Bowman R, Jeffery H, Luben R, et al. A single nucleotide polymorphism in the 3-hydroxy-3-methylglutaryl-coenzyme A reductase gene (*HIMGCR*) influences the serum triacylglycerol relationship with dietary fat and fibre in the European Prospective Investigation into Cancer and Nutrition in Norfolk (EPIC-Norfolk) study. British Journal of Nutrition. 2010;104(5):765-72.

- Srivastava A, Pandey S, Choudhuri G, Mittal B. Role of genetic variant A-204C of cholesterol 7α-hydroxylase (CYP7A1) in susceptibility to gallbladder cancer. Molecular Genetics and Metabolism. 2008;94(1):83-9.
- Hu M, Mak VWL, Chu TTW, Waye MMY, Tomlinson B. Pharmacogenetics of HMG-CoA Reductase Inhibitors: Optimizing the Prevention of Coronary Heart Disease. Current Pharmacogenomics and Personalized Medicine. 2009;7(1):1-26.
- Thompson JF, Man M, Johnson KJ, Wood LS, Lira ME, Lloyd DB, *et al.* An association study of 43 SNPs in 16 candidate genes with atorvastatin response. Pharmacogenomics J. 2005;5:352-8.
- Krauss RM, Mangravite LM, Smith JD, Medina MW, Wang D, Guo X, et al. Variation in the 3-Hydroxyl-3-Methylglutaryl Coenzyme A Reductase Gene Is Associated With Racial Differences in Low-Density Lipoprotein Cholesterol Response to Simvastatin Treatment. Circulation. 2008;117(12):1537-44.
- Chasman DI, Posada D, Subrahmanyan L, Cook NR, Stanton VP, Ridker PM. Pharmacogenetic Study of Statin Therapy and Cholesterol Reduction. JAMA: The Journal of the American Medical Association. 2004;291(23):2821 -7.
- Donnelly LA, Doney AS, Dannfald J, Whitley AL, Lang CC, Morris AD, et al. A paucimorphic variant in the HMG-CoA reductase gene is associated with lipidlowering response to statin treatment in diabetes: a GoDARTS study. Pharmacogenet. Genomics. 2008;18:1021-6.
- Poduri A, Khullar M, Bahl A, Sehrawat B s., Sharma Y, Talwar KK. Common Variants of HMGCR, CETP, APOAI, ABCB1, CYP3A4, and CYP7A1 Genes as Predictors of Lipid-Lowering Response to Atorvastatin Therapy. DNA and Cell Biology. 2010;29(10):629-37.
- Chen Y-C, Chen Y-DI, Li X, Post W, Herrington D, Polak JF, et al. The HMG-CoA reductase gene and lipid and lipoprotein levels: the multi-ethnic study of atherosclerosis. Lipids. 2009;44(8):733-43.
- Freitas RN, Khaw K-T, Wu K, Bowman R, Jeffery H, Luben R, *et al.* HMGCR gene polymorphism is associated with stroke risk in the EPIC-Norfolk study. Eur J Cardiovasc Prev Rehabil. 2010;17(1):89-93.
- Teslovich TM, Musunuru K, Smith AV, Edmondson AC, Stylianou IM, Koseki M, et al. Biological, Clinical, and Population Relevance of 95 Loci for Blood Lipids. Nature. 2010;466(7307):707-13.
- Chien K-L, Wang K-C, Chen Y-C, Chao C-L, Hsu H-C, Chen M-F, *et al.* Common sequence variants in pharmacodynamic and pharmacokinetic pathway-related genes conferring LDL cholesterol response to statins. Pharmacogenomics. 2010;11(3):309-17.
- 30. Cambien F, Tiret L. Genetics of Cardiovascular Diseases. Circulation. 2007;116(15):1714-24.
- Mangravite LM, Thorn CF, Krauss RM. Clinical implications of pharmacogenomics of statin treatment. Pharmacogenomics J. 2006;6(6):360–74.
- Zhu H, Tucker HM, Grear KE, Simpson JF, Manning AK, Cupples LA, et al. A Common Polymorphism Decreases Low-Density Lipoprotein Receptor Exon 12 Splicing Efficiency and Associates with Increased Cholesterol. Hum Mol Genet. 2007;16(14):1765-72.
- Martinelli N, Girelli D, Lunghi B, Pinotti M, Marchetti G, Malerba G, et al. Polymorphisms at LDLR locus may be associated with coronary artery disease through modulation of coagulation factor VIII activity and independently from lipid profile. Blood. 2010;116(25):5688-97.

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