



Anti-Inflammatory and Anti-Oxidant Study of Ethanolic Extract of *Mimosa pudica*

Shofiul Azam^{*1}, Archi Farhana Huda¹, Kishower Shams¹, Prawej Ansari¹,
Mustafa Khalid Mohamed¹, Md Mahmudul Hasan¹, Abul Kalam Azad¹
Kallol Kanti Mondal³ and Shakil Mahmood Zaouad²

^{*1}Department of Pharmaceutical Sciences, North South University, Dhaka, Bangladesh

²Department of Pharmacy, University of Asia Pacific, Dhaka, Bangladesh

³Department of Pharmaceutical Science, Northern University, Dhaka, Bangladesh

ABSTRACT

Objective: Our study includes the investigation of the phytochemical composition and *in vitro* free radical content and anti-inflammatory activity of *Mimosa pudica*. **Materials and Method:** Free radical scavenging assay was done to evaluate the dose dependent reduction of free radical by ethanolic extract of *Mimosa pudica*, this activity was compared with a reference antioxidant i.e. Ascorbic acid. The anti-inflammatory potential of ethanolic extract of *Mimosa pudica* has been determined by using carrageenan-induced paw edema assay and Cotton wool granuloma in rats. **Results:** The IC₅₀ of our sample was 24.55 µg/ml, it's a very much promising result comparing to same reference. At the dose of 300 mg/kg the extract shows considerable inhibitory effect on paw increase 1 hour after carrageenan administration, by inhibiting nearly 50%. The maximum inhibition (43.48%; p<0.001) elicited by the ethanolic extract was recorded after 4 hours after carrageenan injection. Diclofenac sodium which is a reference drug showed a similar inhibitory effect 4 hours after carrageenan administration (50.31%). The cotton wool granuloma is widely used to evaluate the transudative and proliferative components of chronic inflammation. Chronic inflammation occurs by means of the developments of proliferated cells and which can be spread in granuloma form. Non-steroidal anti-inflammatory drugs decrease the size of granuloma which results from cellular reaction by inhibiting granulocyte infiltration, preventing generation of collagen fibers and suppressing mucopolysaccharides. The extract showed significant (p<0.01) anti-inflammatory activity in cotton wool induced granuloma with 33.64% of inhibition at higher dose. **Conclusion:** This finding suggests that ethanolic extract of *M. pudica* possess potent anti-inflammatory activity possibly due to its free radical scavenging properties.

Key words: Anti-oxidant, Carrageenan, Cotton oil, Inflammation, *Mimosa pudica*, Paw edema.

INTRODUCTION

Plants have provided humans with many of their essential needs, including life-saving pharmaceutical agents. Recently the World Health Organization estimated that 80% people worldwide rely on herbal medicines for some aspect. There are more than 270,000 higher plants existing on this planet, but only a few has been explored scientifically for pharmacological use. So, it is anticipated that plants can

Access this article online

Journal Sponsor



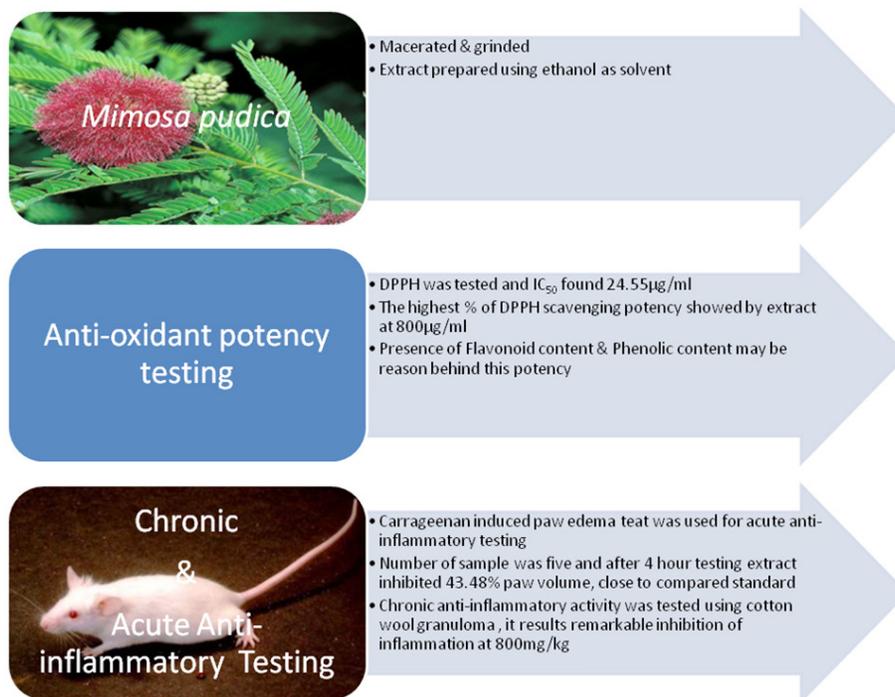
Website:
www.jyoungpharm.org

DOI:
10.5530/jyp.2015.3.14

*Address for correspondence:

Mr. Shofiul Azam, Department of Pharmaceutical Sciences, 27/ka, Kuril, Dhaka-1229, Phone No : +8801815186262, Bangladesh.

E-mail : shofiul_azam@hotmail.com



Graphical Abstract

provide potential bioactive compounds for the development of new 'leads' to combat various diseases. *Mimosa pudica* is most common herb in all locality of Bangladesh and its proved and important medicinal plant. Besides its local and traditional use it has many established pharmacologic use like, ovulation reduction¹, anticonvulsant², antidepressant³, anti-diabetic⁴, antimicrobial⁵, wound healing⁶, snake venom induced hyaluronidase and protease inhibition, snake venom neutralization,⁷ and antioxidant activities.⁸

The inflammatory response involves a complex array of enzyme activation, mediator release, fluid extravasations, cell migration, tissue breakdown and repair which are aimed at host defense and usually activated in most disease condition.⁹ Several phenomena alter the antigenicity of endogenous proteins, including protein denaturation and glycosylation. Protein denaturation may occur during chronic inflammatory phenomena *in vivo* and albumin denaturation was observed in patients with rheumatic diseases and in rats with inflammatory lesions.¹⁰ Lysosomal enzymes released during inflammation have been implied in acute or chronic inflammation. Many of the NSAIDs such as Diclofenac act by inhibiting these lysosomal enzymes or by stabilizing the lysosomal membrane. Since the membrane of RBC is structurally similar to lysosomal membrane, the effect of any substance on stabilization of RBC membrane may be extrapolated to the stabilization of lysosomal membrane^{11,12} It has been reported that leucocyte proteinases play an important role in the development

of tissue damage during inflammatory reactions and significant level of protection was provided by proteinase inhibitors. Hence inhibition of albumin denaturation, RBC membrane stabilization and protease inhibition afford protection against chronic inflammatory conditions.¹³

Reactive Oxygen Species (ROS) play an important role in the pathogenesis of inflammatory diseases. Antioxidants capable of scavenging ROS are expected to improve these disorders.¹⁴ Hence we propose to study the anti-oxidant properties of the extracts for its ferric reducing anti-oxidant power and DPPH radical scavenging activity.

We propose to study anti-inflammatory activity and free radical scavenging capability of *M. pudica* by studying the traditional use on this purpose of this plant. *M. pudica* has been found to use in the treatment of edema, rheumatism, myalgia and in different painful condition in traditional and tribal medicine; however no scientific investigations yet been reported for its anti-inflammatory properties.

The phytochemical study reports (Table 1) following constituents of *M pudica*: alkaloids, steroids, tannins, triterpenes, flavonoids, glycosides, quinines, phenols, saponin, coumarin, c-glycosylflavones.^{2,5} Presence of the flavonoids, phenols etc. encourages us more to study this plant, because these contents are responsible for wide range of anti-oxidant property, that means they aid protection against ROS and reduce damage of cell.

Table 1: Preliminary Phytochemical Testing

Test of Constituent	Intensity of Presence
Alkaloid	+++
Flavonoid	+++
Carbohydrate	+++
Glucoside	+++
Glycoside	+++
Phenol	+++
Saponin	+++
Steroid	+++
Tannin	+++

MATERIALS AND METHODS

Collection and Extraction

The plant was collected from Botanical Garden of Bangladesh, where this plant is harvested widely. It's an annual herb with tiny little cylindrical shaped leaf. We worked by using those leaves. Leaves were separated and dried at room temperature then extracted using cold extraction process; it is so called because no heat is applied in this process. Ethanol was used as solvent; we have got dark greenish black extract weighing 31.8 gm.

Test for Anti-Oxidant

Preparation of solutions

Ascorbic acid was considered for comparative standard, which was concentrated 0.005 gm/ml. Different concentrations for different tests were prepared by serial dilution from this solution, so far. 0.005 gm/ml of plant extract was prepared in ethanolic solvent for stock solution. From this solution, 800 µg/100 µl, 400 µg/100 µl, 200 µg/100 µl, 100 µg/100 µl and 50 µg/100 µl was prepared for further use.

Procedure

In each concentration of plant extract solution, 3 ml of DPPH solution was added and the mixture was incubated for 30 minutes in a dark place for proper reaction. Then the absorbance of the solution was measured at 517 nm using a spectrophotometer (Shimadzu, UV-1601 PC) against blank/control.¹⁴

Anti-inflammatory testing

Animal selection

Healthy male rats were selected weighing between 120-180 g. They were kept at room temperature with 12-h light and dark cycle.

Carrageenan induced paw edema

Carrageenan

Carrageenan (lambda form, FMC Marine Colloids Division, NJ, or type IV, Sigma Aldrich, Poole, UK) was prepared as a 1% W/V solution in 0.9% saline, no more than 24 h before use. Carrageenan powder becomes extremely sticky on contact with water and may form lumps that are difficult to dissolve. Complete solution of solid material is vital to prevent blockage of the hypodermic needle bore and potential injury to the investigator by pressurized ejection of the needle from the syringe or breakage of the syringe barrel in the hand. The lambda form does not gel strongly at room temperature and is injectable to induce an inflammatory response. Inflammation induced by carrageenan, originally described.¹⁴

Procedure

Animals are weighed, randomized into groups (n=5), and kept for 1 week to acclimatize to the laboratory conditions. Test compounds were administered to animals at an appropriate time point before carrageenan injection. Effects of unknown compounds are usually compared to reference compounds whose pharmacology and action in this model are known. Volume of pre-injection paw/paws measured immediately prior to carrageenan injection, Plethysmometre were used to measure the swelling of paw. 100 µL (rat) or 25 µL (mouse) of a 1% solution of lambda carrageenan in 0.9% saline was injected subcutaneously into the plantar region of the left hind paw. Carrageenan injected and control paw volumes were measured hourly as required from 1–6 h and again at 24 h.

Cotton wool granuloma in rats

This method is a chronic anti-inflammatory evaluation model.¹⁵

Induction of Anesthesia

The animals were sedated using proper dose of Ketamin, a general anesthetic used in minor surgery. The anesthetic was given intra-peritoneally and after 10 minutes as anesthesia established, the animal is ready for experiment.

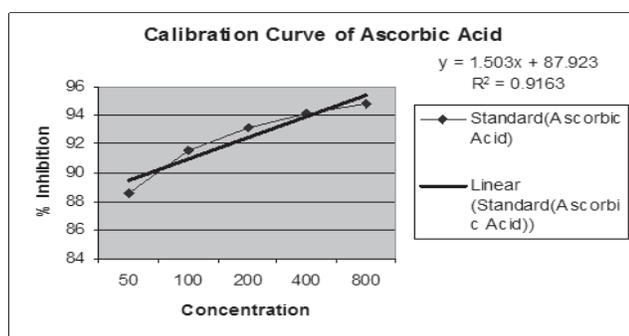
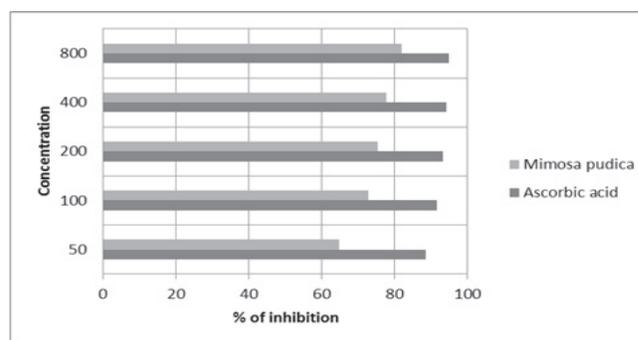
Procedure

The furs of the axilla area were cleaned and were wiped with 70% v/v ethanol. A small subcutaneous incision was made in the axilla region and formed a pouch using blunt ended forceps. 20 ± 1 mg of the sterile cotton pellet was inserted into axilla. The incisions were sutured by sterile catgut/biodegradable surgical stings.

Table 2: Antioxidant Activity of the Ethanolic Extract of *Mimosa pudica*

Sample	Conc. ($\mu\text{g/ml}$)	Absorbance	Absorbance of Control	Mean % DPPH scavenging activity \pm SD	IC ₅₀ V alue ($\mu\text{g/ml}$)
Standard Ascorbic Acid	50	0.080	0.816	88.56 \pm 1.65	1.102 $\mu\text{g/ml}$
	100	0.060		91.56 \pm 0.95	
	200	0.051		93.15 \pm 0.79	
	400	0.046		94.07 \pm 0.26	
	800	0.045		94.82 \pm 0.29	
Ethanol Extract <i>Mimosa pudica</i>	50	0.288		64.72 \pm 0.50	24.55 $\mu\text{g/ml}$
	100	0.214		72.70 \pm 0.98	
	200	0.193		75.39 \pm 1.14	
	400	0.179		77.78 \pm 0.39	
	800	0.134		81.94 \pm 1.53	

All values are Mean \pm SEM, n=3. Independent Sample t-Test followed by Student t-test was performed as the test of significance. P-value was <0.05 which considered statistically significant.

**Figure 1: Calibration curve of ascorbic acid****Figure 2: Percent of DPPH Scavenging Activity of Extracts and Ascorbic Acid**

Removal of cotton

The animals were sacrificed by excess anesthesia on the 8th day and cotton pellets covered by the granuloma tissue were removed surgically. Pellets were separated from extraneous tissue and dried at 60°C until weight become constant. Then the net weight was calculated following the equation:

$$\% \text{ of inhibition} = \frac{W_c - W_d}{W_c} \times 100^{16,17}$$

W_c = Difference in the weight of control group

W_d = Difference in the weight of extract group

RESULT AND DISCUSSION of Free Radical Scavenging of Extarct of *Mimosa pudica*

DPPH was used to evaluate the possible antioxidant principles present in the extract by its radical scavenging capacity measurement (Oyaizu, 1986). The % of DPPH free radical Scavenging was determined by using the following equation.

$$\% \text{ of DPPH free radical Scavenging} = \left[\frac{\text{Absorbance of control} - \text{Absorbance of test sample}}{\text{absorbance of control}} \right] \times 100$$

DPPH radical scavenging is a widely used method to evaluate the free radical scavenging ability of various materials. DPPH is a stable nitrogen-centered free radical, the color of which changes from violet to yellow upon reduction by either the process of hydrogen-or electron-donation. Substances which are able to perform its reaction can be considered as antioxidants and, therefore, radical scavengers. It was found that the radical scavenging activitie of the extract increased with increasing concentration. High total phenol content and total Antioxidant Capacity of the ethanol plant extract may be a reason for its higher DPPH-scavenging activity.

The results were calculated as IC₅₀ values, which denotes the concentration of sample required to scavenge 50% of DPPH free radicals (Table 2).

In this assay, plant extract showed dose dependent scavenging of DPPH radicals (Figure 1 and 2) in a way similar to that of the reference antioxidant ascorbic acid.

Table 3: Paw volume change in different group in Carrageenan induced paw edema testing

Treatment Group	Paw Volume				
	0 hour	1 hour	2 hour	3 hour	4 hour
Control	0.73±0.111	1.19±0.231	1.39±0.071	1.56±0.144	1.61±0.164
Standard	0.76±0.085	0.98±0.058	1.16±0.098	1.05±0.226	0.8±0.094
Drug 100 mg/kg	0.78±0.094	1.05±0.095	1.35±0.156	1.11±0.223	1.07±0.222
Drug 200 mg/kg	0.71±0.069	1.03±0.109	1.29±0.215	1.1±0.227	0.97±0.121
Drug 300 mg/kg	0.75±0.093	1.01±0.241	1.26±0.149	1.06±0.175	0.91±0.265

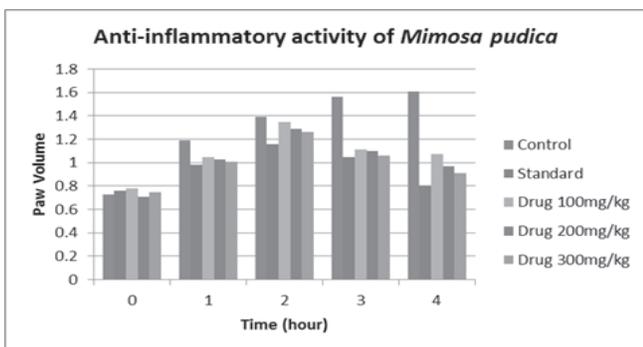
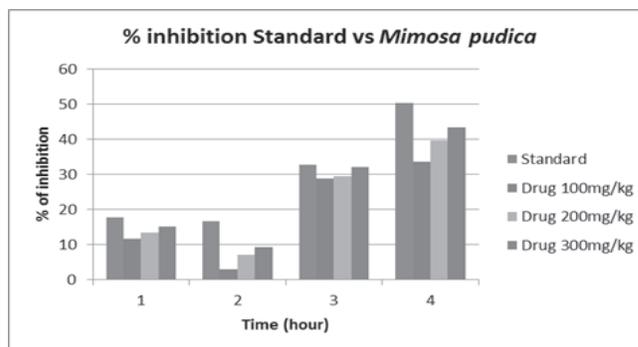
Values are expressed as mean ± standard error of the mean (SEM). Data were analyzed by analysis of variance (ANOVA) followed by post hoc analysis with a one-tailed Dennett's t-test for multiple comparisons.

RESULT AND DISCUSSION of Carrageenan Induced Paw Edema

Table 4: % inhibited by *Mimosa pudica* in comparison with standard in Carrageenan induced paw edema testing

Treated Groups	% Inhibition			
	1 hour	2 hour	3 hour	4 hour
Standard	17.65	16.55	32.69	50.31
Drug 100 mg/kg	11.76	2.88	28.85	33.54
Drug 200 mg/kg	13.45	7.19	29.49	39.75
Drug 300 mg/kg	15.13	9.35	32.05	43.48

For each of the three doses of extract tested (100, 200 and 300 mg/kg), the extract exerted considerable inhibitory effect on paw increase 1 hour after carrageenan administration, with near a 50% inhibition for the dose 300 mg/kg (Table 3 and 4). The maximum inhibition (43.48%, $p < 0.001$) elicited by the ethanolic extract was recorded after 4 hours after carrageenan injection. Diclofenac sodium which is a reference drug showed a similar inhibitory effect (Figure 3 and 4) 4 hours after carrageenan administration (50.31%)

**Figure 3: Anti-inflammatory Activity of *Mimosa pudica*****Figure 4: % Inhibition of *Mimosa pudica* at carrageenan induced paw edema****Table 5: % inhibited by *Mimosa pudica* in cotton wool granuloma test**

Group and Dose of Drug	Weight of wet Cotton Wool	% Inhibition	Weight of Dry Cotton Wool	% Inhibition
Control (Water)	299.17±5.69	-	86.5±0.86	-
Standard (Diclofenac Na 10 mg/ Kg)	159.5±5.76*	46.69	44.33±1.4*	48.75
Dose 1 (Extract 200 mg/Kg)	285.5±2.53*	22.85	85.33±1.69*	23.97
Dose 2 (Extract 400 mg/Kg)	187.11±2.98*	29.12	55.33±1.26*	31.88
Dose 3 (Extract 800 mg/Kg)	180.5±2.85*	31.22	53.33±1.2*	33.64

Values are expressed as Mean ± SEM, n=4 in each group. Data was analyzed by one way ANOVA followed by Dunnett's Test. *P<0.01 as compared to control group, that means statistically significant.

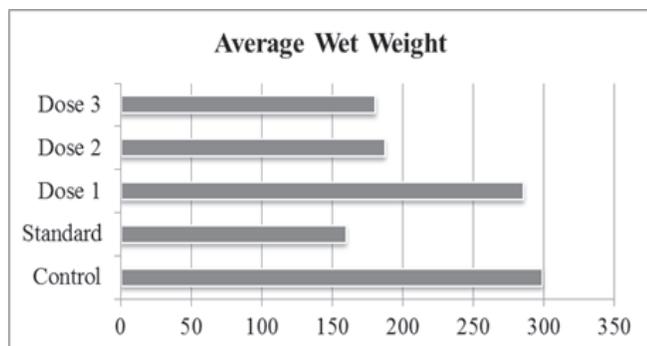


Figure 5: Effect of the Average Wet Weight of cotton Pellets with Respect to different groups

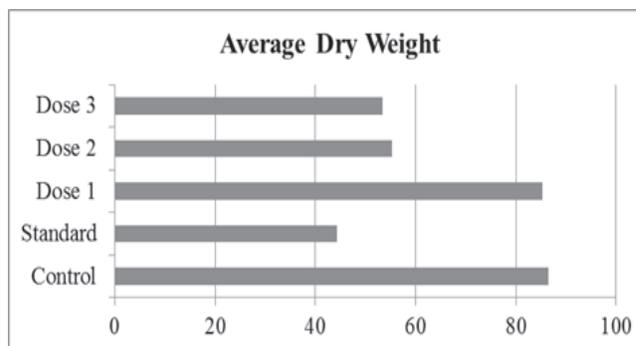


Figure 6: Effect of the Average Dry Weight of cotton Pellets with Respect to different groups

RESULT AND DISCUSSION of Cotton wool granuloma test

The cotton wool granuloma is widely used to evaluate the transudative and proliferative components of chronic inflammation. The moist weight of the wool correlates with transude, the dry weight of the wool correlates with the amount of granulomatous tissues. Chronic inflammation occurs by means of the developments of proliferated cells. These cells can be spread in granuloma form. Non-steroidal anti-inflammatory drugs decrease the size of granuloma which results from cellular reaction by inhibiting granulocyte infiltration, preventing generation of collagen fibers and suppressing mucopolysaccharides. The extract showed significant ($p < 0.01$) anti-inflammatory activity in cotton wool induced granuloma (Table 5) and (Figure 5 and 6) and thus found to be effective in chronic inflammatory condition, which reflected its efficacy in inhibiting the increase in the number of fibroblasts and synthesis of collagen and mucopolysaccharides during granuloma tissue formation.

RESULT of Phytochemical Testing

CONCLUSION

In the present investigation we can conclude that the ethanolic extract of *Mimosa pudica* may have anti-oxidant and anti-inflammatory effect, which supports the traditional use of this plant in various diseases as traditional medicine.

The antioxidant activity of the root extract was tested by DPPH radical scavenging assay. In light of the result of the present study, it can be concluded that the plant extract possesses remarkable antioxidant potential. However, further studies are needed to understand the underlying mechanism of antioxidant action and to isolate the compound(s) responsible for such activity.

It is well known that carrageenan induced paw edema is characterized by biphasic event with involvement of different inflammatory mediators. In the first phase (during the first 2 h after carrageenan injection), chemical mediators such as histamine and serotonin play role, which in second phase (3 h after carrageenan injection) Kinins and prostaglandins are involved. Our results revealed that administration of *Mimosa pudica* extract inhibits inflammation which is caused by chemical or other mediators of inflammation.

The cotton wool granuloma is widely used to evaluate the transudative and proliferative components of chronic inflammation. The moist weight of the wool correlates with transude, the dry weight of the wool correlates with the amount of granulomatous tissues. Chronic inflammation occurs by means of the developments of proliferated cells. These cells can be spread in granuloma form. Non-steroidal anti-inflammatory drugs decrease the size of granuloma which results from cellular reaction by inhibiting granulocyte infiltration, preventing generation of collagen fibers and suppressing mucopolysaccharides. The extract of *Mimosa pudica* showed significant ($p < 0.01$) anti-inflammatory activity in cotton wool induced granuloma and thus found to be effective in chronic inflammatory condition, which reflected its efficacy in inhibiting the increase in the number of fibroblasts and synthesis of collagen and mucopolysaccharides during granuloma tissue formation.

CONFLICTS OF INTEREST

Authors declare no conflict of interest.

ACKNOWLEDGEMENTS

First of all we would like thank North South University

for allowing us to use their lab, unless we will not eligible to finish our work. We also like to give special thanks to icddr'b (international center for diarrheal disease research, Bangladesh) for their help to manage our animal subject. Especial thanks to Mohammad Mamun Ur Rashid, graduate student, Department of Pharmaceutical Sciences, North South University, for his suggestion during article preparation.

Highlights of Paper

- Carrageenan increase cellular metabolism of Kinins and prostaglandins after 3 hr of injection, as well as histamin release increased.
- All these mediators influence inflammation.
- Extract reduces inflammation after 4 hour because it either inhibits the prostaglandin synthesis or kinin.
- Cotton wool induction enhance the cell to proliferate.
- Mucopolysaccharide and collagen fiber generates more, cell become inflamed.
- Extract prevents the inflammation by preventing generation of mucopolysaccharide and collagen fiber, thus cell swelling prevented and chronic inflammation controled.

Author Profile



• Shofiul Azam, is an Graduate student of Department of Pharmaceutical Sciences, North South University. Completed honors (4 year Bachelor degree) from International Islamic University Chittagong. I am hanged up with different plants available in Bangladesh, evaluating their ethnopharmacological property, also trying to identify their phytochemical content. Recently working in a project with another plant and investigating its lipid lowering potency in hypertensive rat model.



• Archi Frahana Huda, Completed her graduation program from North South University and also her 4 years bachelor program from same institute, under the department of Pharmaceutical Sciences. Now she is working in a renowned pharmaceutical farm in Bangladesh (Sk+F Pharmaceutical Ltd.) as a Product Management Officer. She was engaged in different pharmacological property investigation project during her graduate and under-graduate program. Anti-inflammatory and anti-oxidant study of ethanolic extract of *M. pudica*, is also a part of her project.

REFERENCES

1. Valsala S, Karpagaganapathy PR. Effect of *Mimosa pudica* root powder on oestrous cycle and ovulation in cycling female albino rat *Rattus norvegicus*. *Phytother Res*. 2002; 16(2): 190-2.
2. Bum EN, Dawack DL, Schmutz M, Rakotonirina A, Rakotonirina SV, et al. Anticonvulsant activity of *Mimosa pudica* decoction. *Fitoterapia* 2004; 75(3): 309-14.
3. Molina M, Contreas CM, Tellez AP. *Mimosa pudica* may possess antidepressant action in the rat. *Phytomedicine* 1999; 6(5): 319-23.
4. Sutar NG, Sutar UN, Behera BC. Anti-diabetic Activity of the Leaves of *Mimosa pudica* Linn in albino rats. *J Herbal Medicine and Toxicology* 2009; 3(1): 123-6.
5. Gandiraja N, Sriram S, Meena V, Srilakshmi JK, Sasikumar C, Rajeswari R. Phytochemical Screening and Antimicrobial Activity of the Plant Extracts of *Mimosa pudica* L against selected Microbes. *Ethnobotanical Leaflets* 2009; 13(1): 618-24.
6. Kannan S, Jesuraj SAV, Jeeva KES, Saminathan K, Suthakaran R, Ravi KM, et al. Wound healing activity of *Mimosa pudica* Linn formulation. *Inter J Pharmtech Research* 2009; 1(4):1554-8.
7. Girish KS, Mohanakumari HP, Nagaraju S, Vishwanath BS, Kemparaju K. Hyaluronidase and Protease activities from Indian snake venoms: neutralization by *Mimosa pudica* root extract. *Fitoterapia* 2004; 75(3): 378-80.
8. Genest S, Kerr C, Shah A, Rahman MM, Saif GMM, Nigam P, Nahar L, Sarker SD. Comparative bioactivity studies on two *Mimosa species*. *Boletin Latinoamericano y del caribe de plantas Medicinales y Aromaticas* 2008; 7(1): 38-43.
9. Rajendran V, Lakshmi KS. *In vitro* and *in vivo* anti-inflammatory activity of leaves of *symplocos cochinchinesis* (lour) Moore ssp *laurina*. *J Pharmacol* 2008; 3(1): 121-4.
10. Luciano S, Giovanni V, Maria LC, Eleonora M, Laura B, et al. Inhibition of Protein Denaturation by Fatty Acids, Bile salts and other Natural Substances: A New Hypothesis for the Mechanism of Action of Fish Oil in Rheumatic Diseases. *Jpn J Pharmacol* 1999; 79(1): 89-99.
11. Rajurkar R, Jain R, Mataka N, Aswar P, Khadbadi SS. Anti-inflammatory Action of *Abutilon indicum*(L) Sweet Leaves by HRBC Membrane Stabilization. *Res J Pharm Tech*. 2009; 2(2): 415-6.
12. Chatterjee S, Das SN. Anti-arthritis and Anti-inflammatory Effect of a poly-herbal drug its mechanism of action. *Indian J Pharmacology* 1996; 28(2): 166-19.
13. Ilavarassan R, Mallika M, Venkataraman S. Anti-inflammatory and Antioxidant Activities of *Cassia Fistula* Linn bark extracts. *Afr J Trad CAM*. 2005; 2(1): 70-85.
14. Parasuraman S, Petchi RR, Vijaya C, Dhanaraj SA. Evaluation of free radical scavenging properties and hypoglycemic activity of ethanolic extract of *Tridax procumbens* Linn. in Wistar rats. *Drug Dev Ther*. 2014; 5(1): 164-7.
15. Gerhard vogel H, Wolfganga HV, Bernward AS, Jurgin S, Gunter M, Wolfganga FV. Drug discovery and evaluation pharmacological assays; 2nd ed; Berlin, Germany: Spinger; 2002; p-725-71.
16. Intahphuak S, Panthong A, Kanjanapothi D, Taesotikul T, Krachangchaeng C, Reutrakul V. Anti-inflammatory and analgesic activities of *Mallotus spodicarpus* Airy Shaw. *J Ethnopharmacol*. 2004; 90(1): 69-72.
17. Smita S, Shwetha K, Prabhu K, Maradi R, Bairy KL, Shanbhag T. Evaluation of antiinflammatory activity of *Tephrosia purpurea* in rats. *Asian Pac J Trop Med*. 2010; 3(3): 193-5.

ETHICAL CONCERN

Whenever we use any animal model it is our responsibility to take permission from appropriate authority for authentication. We took ethical consent from our university authority and also from icddr'b, when we were collecting animal and using in our study. So the whole study was run in an ethical manner.