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Polyclonal T Lymphocytes Transform into Malignant Cells by Retroviral Mediated Transfer of p21SNFT Gene

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ABSTRACT

Background: p21SNFT (21 kDa small nuclear factor isolated from T cells) gene is known to be increasingly expressed in the cells of Hodgkin disease and Hodgkin cell lines. It was not investigated before if this gene had any effect on polyclonal T lymphocytes. For this purpose, we transduced this gene into polyclonal T lymphocytes and transplanted into mice. Materials and Methods: We used γ -retroviral vector to transduce this gene into polyclonal T lymphocytes, isolated from wild type black-6 mice. After confirmation of p21SNFT expression via marker gene eGFP (enhanced green fluorescent protein) and Western blot analysis, these cells were transplaned into immunocompromised Rag 1 deficient mice. The control group of mice was transplanted with γ -retroviral vector, carrying only marker gene (eGFP). The mice were followed up. The results showed that all 7 mice transplanted with p21SNFT-carrying polyclonal T lymphocytes developed massively enlarged mesenteric lymph nodes after 435 to 550 days of transplantation. Results: Histopathological analyses of these lymph nodes revealed the lymphoma, which resembled human peripheral T cell lymphoma type. Furthermore, immunophenotyping of tumor cells by FACS analysis revealed loss of T cell markers and expression of CD19 marker. From the control group, all five mice remained healthy and did not develop any tumor. Conclusion: The study concludes that γ -retroviral mediated transfer of p21SNFT transforms polyclonal T lymphocytes into malignant cells. **Key words:** γ -retroviral vector, Lymphoma, Malignant transformation, p21SNFT, T lymphocytes.

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INTRODUCTION

The gene *p21SNFT* (21 kDa small nuclear factor isolated from T cells) encodes a basic leucine zipper protein.¹ Through Serial analysis of gene expression and semi-quantitative RT/PCR analyses, this gene was found to be up regulated in Hodgkin disease (HD) and Hodgkin cell lines.^{2,3} Since this gene was upregulated in Hodgkin disease cells, we wanted to further investigate if *p21SNFT* could have oncogenic potential for lymphocytes. In the current study we selected wild type polyclonal T lymphocytes. T lymphocytes, which are produced by hematopoietic stem cells in the bone marrow, migrate to the thymus for further processing of selection and maturation. Afterwards, they leave thymus and enter the blood circulation and lymphatic system.^{4,5} T lymphocytes get their stimulatory niche through their receptors TCRs (T cell receptors), by interacting with molecule spMHC (self-peptide major histocompatibility complex) expressed on antigen presenting cells. In a human or a wild type mouse, these lymphocytes express different types of TCRs, hence are polyclonal. This polyclonality provides the lymphocytes with a variety of TCR expressing T lymphocytes, which interact and fight a variety of pathogens. Thus, T lymphocyte diversity provides us with a powerful defense system against a variety of pathogens.^{6,7} In the current experimental setup, we chose T lymphocytes for two reasons. First, these cells have been used in gene therapy.8 Therefore, we wanted to further confirm the safety issue of these cells in gene therapy. Second, as p21SNFT gene was found to be upregulated in Hodgkin disease cells and Hodgkin disease has been investigated to originate from B⁹ and T lymphocytes.¹⁰ Hence, we wanted to investigate its oncogenic potential in polyclonal T lymphocytes.

We used Rag 1 deficient mice as the recipients of *p21SNFT* transduced T lymphocytes. These mice are immunodeficient as they do not have mature T and B lymphocytes, hence easily accept the transplanted cells as there would be no immune rejection.

For the transfer of *p21SNFT* gene into polyclonal T lymphocytes, we used γ -retroviral vector. This vector is derived from enveloped RNA virus. The vector carries the desired gene into the target cell where it reverse-transcribes to DNA and integrates into the host genome. These vectors are already reported to support a long term expression of the transgene.¹¹ Therefore, we used γ -retroviral vector to transfer *p21SNFT* gene. Hence, by using this setup, we could transduce *p21SNFT* effectively with its full expression on protein level in polyclonal T lymphocytes and investigate its effects.

MATERIAL AND METHODS

Gammaretroviral vector and packaging of p21snft gene

A γ -retroviral vector MP91/eGFP was used to transfer *p21SNFT* gene into polyclonal T lymphocytes. Empty MP91/eGFP vector was used as a control as described.¹² We purchased the cDNA of human *p21SNFT* gene from GeneArt Company and cloned it into MP91/eGFP vector. This vector codes for eGFP (enhanced green fluorescent protein), which acts as a marker gene as it fluoresces after it is expressed in the cells. The gene *p21SNFT* was cloned in MP91/eGFP vector as described.¹²

Figure 1 shows the control γ -retroviral vector 'MP91/eGFP' and *p21SNFT*-cloned γ -retroviral vector 'MP91/p21SNFT'.

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Figure 1: Molecular map of γ -retroviral vector. MP91/p21SNFT vector (upper). Control vector MP91/eGFP (lower). p21SNFT gene was cloned into multiple cloning site (mcs) of the control vector. IRES: Internal ribosomal entry site; eGFP: Enhanced green fluorescent protein; wPRE: Woodchuck hepatitis virus posttranscriptional regulatory element; LTR: Long terminal repeat.

The MP91/eGFP and MP91/p21SNFT vectors were packaged by calcium phosphate mediated transient transfection of 293 T cells as described.¹² The supernatant, containing the viral particles (eGFP / *p21SNFT*) were filtered through the strainer and used to transduce polyclonal T lymphocytes.

Cell culture and animal experimental work

Animal experiments were approved by the Regional Council Darmstadt. For animal experiments, mice were purchased from Charles River Laboratories (Sulzfeld, Germany) and The Jackson Laboratory (Bar Harbor, USA).

Polyclonal T lymphocytes were isolated from six to eight weeks old wild type black 6 mice. These donor mice were first anesthetized with isofluran and then sacrificed by dislocating the cervical spines. 70% ethanol was sprayed over the skin to disinfect the abdominal and chest parts of the sacrificed mice. After careful dissection, spleen and mesenteric lymph nodes were excised and homogenized with 1% PBS buffer. The cells were filtered through cell strainer and washed with 1% PBS buffer. Isolated cells were stimulated with anti CD3/CD28 antibodies-coupled paramagnetic beads for four days. The cells were analyzed by using FACS (fluorescent activated cell sorting) to confirm the expression of T cell markers CD3 and TCR α/β .

Afterwards, these polyclonal T lymphocytes were transduced with the MP91/p21SNFT and control (MP91/eGFP) supernatants. The transgene expression, represented by eGFP expression, was measured after 36 hr by FACS. Further confirmation of *p21SNFT* gene on protein level expression was done by performing Western blot. The cells were then washed with 1% PBS buffer and transplanted into RAG 1 deficient recipient mice. $1-2\times10^7$ cells / mouse were injected into tail vein. 7 mice were transplanted with p21SNFT transduced polyclonal T lymphocytes and 5 mice were transplanted with the eGFP transduced polyclonal T lymphocytes which served as the control group. After follow up of ~12 months, the mice were sacrificed due to limited life expectancy and the cells were isolated from spleen and mesenteric lymph nodes. After homogenizing and washing with 1% PBS buffer and confirming by FACS analysis for eGFP expression, the cells were serially transplanted into 6 to 8 weeks old RAG 1 deficient mice. The protocol was followed as described.^{12,13}

Blood analysis

Blood was withdrawn from lateral tail vein by making a small incision with a scalpel during the experimental time, or directly by puncturing the heart after sacrificing the mice. 50-100 μ l of blood was collected in an EDTA coated capillary (Microvette) for FACS and hemogram analyses. Hemogram was performed using Scil vet animal blood counter (Scil Animal Care, Viernheim, Germany).

Histopathological analysis

Mice with the signs of sickness, or enlarged abdomen were sacrificed by cervical dislocation. Spleen, liver, lymph nodes and bone marrow were isolated and fixed in Zinc Formal Fixx (Thermo Shandon, Pittsburgh, USA). After 24 hours, the Zinc Formal Fixx was exchanged with 70% ethanol. Histological sections and diagnoses were done by Pathology Institute at University of Frankfurt.

FACS analysis

~5×10⁵ cells from each sample (blood / tumor mass) were used. Each sample was measured in triplicate. The tumor mass was homogenized with the stamp of a syringe through a cell strainer in a 6-well plate and put in a 5 ml polystyrol FACS tube. The cells were washed with 1 × PBS buffer and suspended in 200 µl PBS. 1 µl of each anti-mouse antibody for CD3, TCR α/β , CD19, CD45.1 and CD45.2 markers (Invitrogen) were added. After incubating the cells for 15 min. in dark, Cal-lysis solution (Invitrogen) was added to the samples to lyse red blood cells (RBCs). The cells were washed three times in 3 ml of 1 × PBS buffer by centrifuging at 1400 rpm / 3 min. The supernatant was discarded and the cells were resuspended in 400 µl of 1 × PBS buffer. The samples were analyzed using FACSCalibur and Cellquest pro software.

Western blotting

Western blotting was performed for the confirmation of expression of p21SNFT on protein level. $3x10^6$ tumor cells were washed three times with 3 ml cold sterile 1 x PBS buffer by centrifuging for 2 min at 13000 rpm. The cell pellets were resuspended in ice-cold lysis buffer and vortexed and incubated for 15 min. on ice. The lysates were then spinned down for 10 min. at 13000 rpm in a microcentrifuge (4°C) to settle the cell debris down. Proteins in the supernatant were harvested and transferred to a 1.5 ml reaction tube. Mouse anti *p21SNFT* antibodies (Abnova) were used for blotting. The detailed protocol was followed as described.¹²

RESULTS

All 7 mice, transplanted with *p21SNFT*-transduced polyclonal T lymphocytes developed lymphoma after 435 to 550 days of transplantation. Figure 2 shows the survival curve.



Figure 2: Survival curve of mice. All mice from p21SNFT group developed lymphoma (red line) after 435 to 550 days of transplantation. None of the mice from control group (green line) developed any malignancy.

The mice developed enlarged belly and loss of weight. Mice were sacrificed and immediately blood samples were collected from the heart with the help of a syringe. Blood complete pictures were performed, which did not show any abnormality. For further examination, autopsies of the mice were performed, which showed massively enlarged mesenteric lymph nodes (Figure 3a). The histopathological analyses of these masses showed diffusely spread intermediate sized mature lymphocytes with clear cytoplasm and oval nucleus, comprising the features of peripheral T cell lymphoma (Figure 3b).

For further investigation, FACS analysis was performed to determine the phenotype of the tumor cells. All the tumors showed similar phenotypes. All eGFP expressing cells were positive for CD19 and negative for CD3 and TCR α/β (Figure 4). For further confirmation, Western blotting was performed, which showed the expression of *p21SNFT* on protein level in tumor cells (Figure 5).



Figure 3: (a) Gross features of tumor mass. Grossly enlarged mesenteric lymph node. (b) Histopathological features of enlarged lymph node. Diffusely spread intermediate sized mature lymphocytes with clear cytoplasm and oval nucleus, comprising the features of peripheral T cell lymphoma (H&E staining, magnification X40).



Figure 4: FACS analysis of enlarged lymph nodes. Almost all eGFP expressing cells were positive for CD19 and negative for CD3 TCR α and TCR β .



Figure 5: Western blot analysis. Tumor cells expressed p21SNFT protein (14.5kDa). Control cells did not express this protein.

DISCUSSION

In this study, we showed that retrovirus mediated transfer of *p21SNFT* gene into polyclonal T lymphocytes and subsequent transplantation of these cells into mice induced lymphoid tumors of mature type, which resembled human peripheral T cell lymphoma. There are three central findings of our research work which support the causative role of *p21SNFT* in the development of lymphoma. First, all the tumor cells expressed the eGFP marker gene, which was confirmed by FACS analysis. Second, the tumor cells expressed *p21SNFT* gene on protein level, which was confirmed by performing Western blot analysis. Third, the tumors originated from the donor cell compartment, not from the recipient mice themselves, which was confirmed by using CD45.1 and CD45.2 markers. The donor cells were positive for CD45.1 and the recipients for CD45.2 marker. Hence, all eGFP expressing tumor cells were positive for CD45.1, which indicate that the tumors were induced in the *p21SNFT* transduced T lymphocytes.

For further confirmation of the phenotype of tumor cells, we performed CD marker analysis, which revealed the loss of T cell markers (CD3 and TCR). Several studies have shown the loss of T cell markers and aberrant expression of CD markers by T cell lymphomas. A study on hepatosplenic T cell lymphoma showed that the tumor cells lost the expression of CD3.¹⁴ Another study also revealed loss of TCR marker in cutaneous T cell lymphoma.¹⁵ In our experiments, the tumor cells lost T cell markers and expressed CD19, which is a B cell marker. The aberrant expression of CD19 by T cell lymphoma cells was already described by several researchers.¹⁶⁻¹⁸

Another study reported a case of peripheral T-cell lymphoma, in which the cells not only expressed CD19, but also other B cell markers such as CD20 and CD79a.¹⁹

The tumors in our experimental mice developed after a longer latency period, which resemble some of the human lymphomas which develop after a longer latency period of years.²⁰ The longer latency period indicates that in addition to the *p21SNFT* gene, there may have occurred other additional mutations which allowed the outgrowth of a transformed tumorigenic clone, resulting in the tumor mass with identical cell morphology. The longer latency period for the development of lymphoma have already been described.²¹ Hence, in this regard several cooperating genes may be involved in the tumorignesis in our experimental mice which need to be further explored by performing DNA microarray analysis. A recent study on genetic analysis of peripheral T cell lymphoma revealed expression profile of 155 genes which were differentially expressed in comparison to the normal T lymphocytes. Among those 155 genes, 64 were upregualted and 91 were downregulated. Majority of the upregulated genes were known to play a role in transcription, proliferation, cell adhesion and matrix remodeling, while many of the downregulated genes were known to play a role in apoptosis. All these events contribute to the development of lymphoma.22

In the current study, we used γ -retroviral vector to transfer *p21SNFT* into T lymphocytes. Although we used the control group of mice, transplanted with empty γ -retroviral vectors, from which none of the mice developed any malignancy, it might still be possible that T lymphocytes got transformed into malignant cells as a result of insertional mutagenesis by γ -retroviral vectors. Retroviral insertional mutagenesis, resulting in cancerous transformation of normal cells have already been previous-ly reported.^{23,24} As the mature T lymphocytes could be transformed into malignant cells in our experimental set up, the safety issue of T lymphocytes in gene therapy should further be investigated.

Since we used limited number of mice in our experiments, further experiments on *p21SNFT* with increased number of mice with more

14.5 kDa

molecular analyses need to be carried out to better understand its pathogenic role in the development of lymphoma.

CONCLUSION

The results of our present study show that γ -retroviral mediated transfer of *p21SNFT* causes malignant transformation of polyclonal T lymphocytes.

ACKNOWLEDGEMENT

This study was funded by Deutsche Forschungsgemeinschaft, for which we are thankful to it.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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Article History: Submission Date :03-02-2020 ; Revised Date : 15-02-2020; Acceptance Date : 17-03-2020. Cite this article: Kumar A, Sengodan B, Lal C, Leela A, Fernandez K. Polyclonal T Lymphocytes Transform into Malignant Cells by Retroviral Mediated Transfer of *p21SNFT* Gene. J Young Pharm. 2020;12(1):71-4.